Anti-angiogenic peptides identified in thrombospondin type I domains

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Abstract

Thrombospondin 1, the prototypical protein of the thrombospondin protein family, is a potent endogenous inhibitor of angiogenesis. Although the effects of the thrombospondin 1 on neovascularization have been well studied, little is known about the anti-angiogenic potency of other proteins or peptide fragments derived from the proteins in this family. Here we identify a set of 18 novel, anti-angiogenic 17- to 20-amino acid peptides that are derived from proteins containing type I thrombospondin motifs. We have named these peptides adamtsostatin-4, adamtsostatin-16, adamtsostatin-18, cartilostatin-1, cartilostatin-2, fibulostatin-6.2, fibulostatin-6.3, papilostatin-1, papilostatin-2, properdistatin, scospondistatin, semastatin-5A.1, semastatin-5A.2, semastatin-5B, thrombostatin containing-1, thrombostatin containing-3, thrombostatin containing-6, and wispostatin-1 to reflect their origin. We further demonstrate that these peptides inhibit the proliferation and migration of human umbilical vein endothelial cells in vitro. The anti-proliferative and anti-migratory properties of the identified peptides may be important in maintaining angiogenic homeostasis in vivo and make these peptides suitable candidates for use as anti-angiogenic pharmaceutical agents in numerous therapeutic applications.

Keywords: Angiogenesis; Endogenous; Inhibitor; Proliferation; Migration; Endothelial cell

The thrombospondin family of proteins consists of a group of five prototypical members (TSP-1, -2, -3, -4, -5) that contain a number of modules, among which are a globular amino terminal motif, a pro-collagen homology region, three type I thrombospondin repeats, three EGF or type 2 repeats, and a globular carboxy-terminal region [1]. Two of the five members of this family, thrombospondin 1 (TSP-1) and thrombospondin 2 (TSP-2), have the highest degree of similarity, both in terms of amino acid identity and structural organization.

The thrombospondins are known to have anti-angiogenic properties [2–4]. Thrombospondin 1, the first identified endogenous inhibitor of angiogenesis, has been shown to play a critical role in inhibiting neovascularization, and therefore tumor growth and metastasis. Thrombospondin 1 inhibits the proliferation and migration of endothelial cells both in vitro and in vivo [5].

A disintegrin and metalloproteinases (ADAMs) are a family of proteins with sequence similarity to the repolysin family of snake venoms. These proteins share the metalloproteinase domain with matrix metalloproteinases (MMPs). They are structurally classified into two groups: the membrane-anchored ADAM group and the ADAM with thrombospondin motifs (ADAMTS) group. These molecules are involved in a variety of biological events, including cell adhesion, cell fusion, cell migration, membrane protein shedding, and proteolysis [6]. Like the thrombospondins, the ADAMTS proteins are modular and contain a metalloproteinase catalytic domain, with a repolysin-like zinc binding motif; a disintegrin-like domain; and a type I thrombospondin repeat. This type I TSP repeat is similar to the type I repeats found in TSP-1 and TSP-2.

Peptides from the type I thrombospondin domains of ADAMTS-1 (METH-I) and ADAMTS-8 (METH-II) have been shown to be anti-angiogenic [7,8]. Both can inhibit
vascular endothelial growth factor (VEGF)-induced angiogenesis in the chick chorioallantoic membrane assay as well as fibroblast growth factor (FGF-2)-induced neovascularization in the corneal micropocket assay [7,8].

By using a bioinformatics algorithm, we have recently identified a set of 18 peptides of 17–20 amino acids that are similar to the type I thrombospondin domains of the aforementioned endogenous inhibitors of angiogenesis. These novel peptides are derived from the associated type I thrombospondin repeats of human endogenous proteins and share similarities to the known inhibitors. Several of the identified short peptides are derived from one of several ADAMTS proteins whose anti-angiogenic potency has not been identified, or from an extracellular matrix-residing protein, such as the cartilage intermediate layer protein (CILP), a fibulin, or papillin. We now present experimental evidence demonstrating that peptides derived from the type I thrombospondin repeats of these proteins are anti-angiogenic, inhibiting the proliferation and migration of human umbilical vein endothelial cells (HUVECs) in vitro.

Materials and methods

Cell culture. HUVECs from a single donor were obtained from Cambrex (Walkersville, MD, USA). The cells were propagated in EGM-2 medium, consisting of a basal cell medium with 2% FBS, growth factors (bFGF and VEGF) and antibiotics (gentamicin/amphotericin B). All the cells used were from passage 3 to 6.

Peptide synthesis and handling. The peptides were produced using a solid-phase synthesis technique by a commercial provider (Abgent, San Diego, CA, USA). HPLC and mass spectroscopy analyses of each peptide were performed. For each of the peptides the synthetic procedure yielded 10 mg of >95% purity. The peptides were provided in solid form and solubilized in water before use. The molecular weight of each peptide was confirmed by mass spectrometry. In the case of highly hydrophobic peptides, dimethylsulfoxide (DMSO) at a maximum concentration of 0.1% (v/v) was used as a solvent. We experimentally verified that at this concentration the solvent had no effect on the experimental results.

Results and discussion

Using a bioinformatics analysis, we have identified a set of 18 peptides derived from type I thrombospondin repeats of different proteins that show similarity to known angiogenesis inhibitors derived from these repeats, such as METH-I (ADAMTS-1) and METH-II (ADAMTS-8) and the thrombospondin repeats of the thrombospondin 1 protein (Table 1). Here, we first describe the parent proteins and then present the results of our proliferation and migration experiments. We then summarize the data demonstrat-
ing that all the identified peptides can inhibit the proliferation and migration of human umbilical endothelial cells in vitro. We refer to these peptides as anti-angiogenic or exhibiting anti-angiogenic activity, with a realization that subsequent in vivo validation of these properties in different tissues is necessary.

As already mentioned, the ADAMTS family of proteins has been previously shown to contain two members, ADAMTS-1 and ADAMTS-8, with anti-angiogenic potency. Here we have identified three novel peptides derived from different ADAMTS proteins: one from ADAMTS-4, which we called adamtsostatin-4; one from ADAMTS-16, which we called adamtsostatin-16; and one from ADAMTS-18, which we called adamtsostatin-18. Cartilage intermediate layer proteins (CILPs) are glycoproteins that are found in the interterritorial matrix of the deeper layers of cartilage and are more prevalent in older tissues. Here we have identified two small peptide fragments derived from CILP-1 and CILP-2, which we have named cartilostatin-1 and cartilostatin-2, respectively.

Fibulins are extracellular matrix secreted glycoproteins that have been shown to modulate cell morphology, growth, adhesion, and motility during tumor progression. We have identified two anti-angiogenic peptides derived from the TSP-1 repeat of fibulin-6. We have called these peptides fibulostatin-6.2 and fibulostatin-6.3. Papilin is a proteoglycan-like sulfated glycoprotein. Here we have identified two anti-angiogenic peptides from papilin, named papilostatin-1 and papilostatin-2, which are derived from its TSP-1 repeat. Properdin (factor P) is a plasma protein that is active in the alternative complement pathway of the innate immune system. The anti-angiogenic peptide derived from properdin has been named properdistatin.

Different members of the semaphorin protein family induce neural guidance cues and also participate in developmental and postnatal vessel formation and patterning. Here, we have identified three peptides derived from the type I thrombospondin repeats of semaphorin 5A, named semastatin-5A.1 and semastatin-5A.2, and from semaphorin 5B, named semastatin-5B, with anti-angiogenic properties. SCO-spondin is a large glycoprotein involved in axonal pathfinding. Here, we show that a peptide derived from a TSP-1 repeat of SCO-spondin, which we call scospondistatin, also inhibits the proliferation and migration of endothelial cells.

Fig. 1. Effect of the 18 peptides derived from the type I thrombospondin repeat-containing proteins on the proliferation of HUVECs. After 3 days of incubation with the test peptides, the endothelial cells were detected using a WST-1 colorimetric assay and counted. The results are scaled so that 0% represents the optical signal from the negative control (endothelial cells incubated with medium containing growth factor and serum, data not shown), and 100% represents the signal from the positive control (cells incubated with 100 ng/ml TNP-470, data not shown). Vertical bars indicate the standard error. All values are significantly different from 0% at \( p < 0.001 \), except those marked by NS (non-significant). In all cases, the standard error for the controls was <3% (\( n = 8 \)).
We have also identified three peptides from thrombospondin type I domain-containing proteins -1, -3, and -6 (TSRC1 or ADAMTS-like 4, TSRC3 and TSRC6). These are transmembrane proteins that contain TSP-1 repeats. The three identified peptides, which we have named thrombostatin containing-1, -3, and -6, also inhibit the proliferation and migration of endothelial cells. The last identified peptide is derived from the type I thrombospondin repeat of WNT1 inducible signaling pathway protein 1 (WISP-1). WISP-1 is the fourth member of the CCN family (CCN-4). Wispostatin-1, the peptide we have identified within the type I thrombospondin domain of WISP-1, also exhibits anti-angiogenic activity.

The identified peptides inhibit the proliferation of HUVECs in vitro

A significant determinant of the peptides’ ability to inhibit angiogenesis is their activity in suppressing the proliferation of endothelial cells. Therefore, we tested the ability of the peptides to suppress the proliferation of HUVECs in vitro. The results from the proliferation assay were scaled to allow us to express the peptide activity relative to the positive control, 100 ng/ml of TNP-470, and are presented in Fig. 1.

From the results in the proliferation assay for the 18 tested peptides we identified two patterns of behavior. A group of peptides, composed of cartilostatin-2, fibulostatin-6.3, properdistatin, and scospondistatin, showed a monotonic dose response, with the anti-proliferative activity increasing with increasing the peptide concentration. In this case, saturation was reached with increasing the peptide concentration. Within this group of peptides, we identified a subset of peptides that exhibited a monotonic dose response, but their activity had already reached saturation at the minimum tested concentration of 0.01 µg/ml. In this subset were adamtsostatin-16, adamtsostatin-18, and semastatin-5B. A second group of peptides, adamtsostatin-4, cartilostatin-1, fibulostatin-6.2, papilostatin-2, semastatin-5A.1, semastatin-5A.2, thrombostatin containing-3, and wispostatin-1, was characterized by a biphasic dose response. The peptide activity was non-monotonic, increasing with increasing peptide concentration, reaching a maximum, and then declining with increasing the peptide concentration. This non-monotonic activity is typical of endogenous anti-angiogenic peptides that have been previously described. Examples of such biphasic responses

![Fig. 2. Effect of the peptides on the migration of HUVECs in a modified Boyden chamber migration assay.](https://example.com/fig2.png)

Endothelial cells were allowed to migrate for 20 h in the presence of 20 ng/ml VEGF and 30 µg/ml peptide solution, then stained with calcine and counted. The fluorescent signal was initially scaled so that 0% represents the negative control (endothelial cells in serum- and growth factor-free medium) and 100% the positive control (migration in the presence of 20 ng/ml VEGF). The percentage migration inhibition, shown in the figure, was calculated. Vertical bars indicate the standard error. All values are significantly different from 0% at \( p < 0.001 \), except those marked by NS (non-significant). In all cases, the standard error for the controls was <5% (\( n = 8 \)).
include the activity exhibited in proliferation experiments by full-length endostatin [12,13] and its small-fragment derivatives [14]. Such biphasic dose-response activity is also exemplified by peptides derived from previously discovered anti-angiogenic fragments of thrombospondins: for example, the anti-angiogenic fragments derived from the TSP-1 [15] and TSP-2 [16] proteins and anti-angiogenic fragments derived from SPARC [17,18] and urokinase [19].

In order to exclude false-positive results that could be attributed to the nature of the peptides tested we repeated the proliferation experiments using two scambled peptides, where the amino acid sequences of the peptides were randomly permuted. The results obtained in the proliferation experiments using the scrambled peptides were not statistically different than those for the negative control (data not shown).

Among the ADAMTS-derived peptides, adamtsostatin-4 exhibited a biphasic response, with a maximum activity of 30% at 10 μg/ml, while adamtsostatin-16 and adamtsostatin-18 had already reached saturation at the lowest concentration tested (0.01 μg/ml), with 15% and 30% activity, respectively. Cartilostatin-1 showed a biphasic response, with a maximum activity of 30% at 0.1 μg/ml, while cartilostatin-2 was monotonic, with a maximum activity 50% at the highest concentration tested. Fibulostatin-6.2 had only a non-significant level of activity in the proliferation assay, with a maximum activity of 10%, while fibulostatin-6.3 was monotonic, with a maximum activity of 40%. Papilostatin-1 had only minimal activity in the proliferation assay, but papilostatin-2 was biphasic, with a maximum activity of 40% at 0.1 μg/ml. Properdistatin reached saturation at 20% activity at 30 μg/ml, and similarly scospondistatin also showed a maximum activity of 20% at 30 μg/ml. Of the semaphorin-5 proteins, the peptides derived from semaphorin-5A were biphasic, and both reached maximal activity (40% for semastatin-5A.1 and 30% for semastatin-5A.2) at 0.1 μg/ml. The activity of semastatin-5B reached saturation at 25% activity at the lowest concentration tested. Among the thrombospondin type I domain-containing proteins, thrombostatin containing-1 and -6 were potent inhibitors, completely suppressing the migration of the endothelial cells at 30 μg/ml. Thrombostatin containing-1 activity was actually saturated at 10 μg/ml, while at the same concentration thrombostatin containing-6 exhibited 70% inhibition of migration (Fig. 2).

In summary, in the present study we have identified a set of 18 novel peptides of 17–20 amino acids that are derived from proteins containing type I thrombospondin repeats, and we provide evidence that these 18 peptides have anti-angiogenic activity, inhibiting the proliferation and migration of HUVECs in vitro. In our proliferation assay, we identified two distinct populations of peptides on the basis of their activity profiles: a set that exhibited a monotonic dose response, showing increasing activity with increasing peptide concentration, and a set that exhibited a non-monotonic, biphasic dose response. In the second set, the peptide activity reached a maximum and then decreased with increasing its concentration.

Recent studies concerning the mechanisms of the anti-angiogenic activity of TSP-1, the prototype type I thrombospondin repeat-containing protein, have implicated CD36 as the cell-surface receptor that mediates its effects on endothelial cells [20]. CD36 is an 88-kDa transmembrane glycoprotein expressed on endothelial cells and a collagen-binding molecule. Analyses using affinity chromatography have demonstrated that various motifs of the type I thrombospondin repeats of the TSP-1 protein can bind to CD36 [21]. Recently, β1 integrins were identified as critical components of the thrombospondin repeats’
anti-migratory effects on endothelial cells [22,23]. The epitope that is responsible for the α3β1 integrin binding maps within the amino-terminus of the thrombospondin-1 protein [24]. Future studies, including mutagenesis studies, need to be performed with the identified peptides in order to determine whether this receptor interaction can explain any or all of the data we present here.

Angiogenesis is regulated by the orchestrated expression of endogenous regulatory elements [25–29], which include an array of growth factors that stimulate and control the proliferation and migration of endothelial cells. There is also a growing population of endogenous regulators that work competitively by inhibiting both of these processes, thus suppressing angiogenesis. The fine control of these two opposing elements, referred to as the angiogenic balance [30], is vital for the homeostasis of a physiologic tissue and is disrupted during pathologic conditions, such as cancer. Thus, the identification of novel endogenous anti-angiogenic peptides that may play a role in both physiologic and pathological conditions, such as the peptides described here, have the potential to increase our understanding of angiogenesis in health and disease.

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