**Drying down lipids**

**Updated November 2011**

1. Wash vials
	* Wash flat-bottom glass vials with soap and then rinse well with water.
	* Rinse with 10% HCl and then rinse really well with water.
	* Tap out excess water then place on hot plate (open end up) to dry.
	* Label vials with lipid and amount.
2. Measure out lipids
	* Our lipid stocks are typically 25 mg/ml with 100 mg per vial (so 4 ml per vial).
	* Calculate what volume you want for however many mgs (use the whole bottle up).
	* Measure this out into vials using Hamilton syringe **(non-cemented!!)**.
	* Last vial will have slightly less, note on vial.
	* Clean syringe with methanol and water.
	* Evaporate off chloroform by placing vials in drying thingy in hood.
		+ - * Unhook tube and turn on nitrogen, adjust pressure to 2 psi.
				* Hook tube to machine and lower nozzles into vials.
				* Turn on heat and let dry for ~5 min.
3. Place lipids under vacuum overnight
	* Cover vials in parafilm and poke holes in top
	* Place vials in tray and place in speed vac (have to remove rotor). Turn on vacuum and leave overnight (alternatively, can place vials in jar and hook up to freeze dryer overnight).
	* In morning, remove vials from speed vac and blow nitrogen into each vial before capping.
	* Cap vials, wrap with parafilm, and store in -20 freezer.